A New Cembranoid Diterpene and Other Related Metabolites from the South-China-Sea Soft Coral *Lobophytum crassum*

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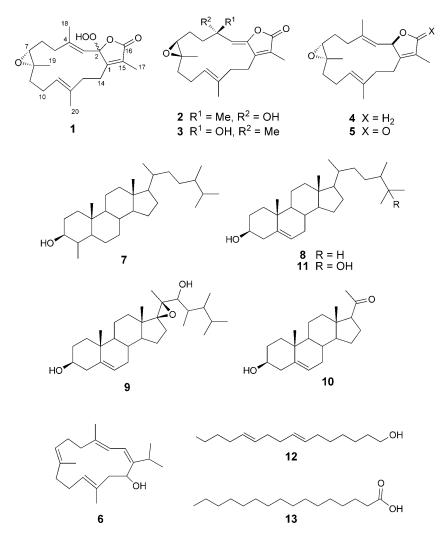
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A new hydroperoxy-substituted cembranoid diterpene, 2-hydroperoxysarcophine (=($1aR^*,4E,11-E,14aR^*$)-2,3,6,7,10a,13,14,14a-octahydro-10a-hydroperoxy-1a,5,8,12-tetramethyloxireno[9,10]cyclote-tradeca[1,2-b]furan-9(1aH)-one; **1**), was isolated from the marine soft coral *Lobophytum crassum*. Also isolated were two other cembranoid diterpenes, obtained for the first time from a natural source, *i.e.*, $7\beta,8\beta$ -epoxy-4 α -hydroxycembra-1(15),2,11-trien-16,2-olide (**2**) and $7\beta,8\beta$ -epoxy-4 β -hydroxycembra-1(15),2,11-trien-16,2-olide (**3**), along with three further cembranoid derivatives, five sterols, and two open-chain metabolites. Their structures and relative configurations were elucidated on the basis of extensive spectroscopic analyses including 1D- and 2D-NMR, and HR-ESI-MS experiments.

Introduction. – A large number of highly functionalized cembranoid diterpenes, sterols, and other related metabolites have been isolated and identified from marine soft corals, especially from the genera *Lobophytum*, *Sarcophyton*, and *Sinularia*, all of which belong to the family Alcyoniidae within the order of Alcyonacea [1][2]. *Lobophytum crassum* is a common soft-coral species widespread in Indo-Pacific reefs [3]. This species has been reported to contain cembranoid diterpenes [4–7], eudesmane-based diterpenoids [7], polyhydroxylated sterols [2][8], and other related compounds [9]. A literature survey revealed that, so far, the reported new cembranoid diterpenes of *L. crassum* origin can be classified into two types: compounds with or without a lactone ring. Interestingly, all new cembranoid diterpenes so far isolated from this species that possess such a lactone ring also have a $\Delta^{15,17}$ unsaturation.

As part of our studies on secondary metabolites from marine organisms from the Chinese Sea, we wish to report in this paper the isolation and structure elucidation of a new cembranoid diterpene, 2-hydroperoxysarcophine (1), along with the two cembranoid diterpenes $7\beta_8\beta$ -epoxy- 4α -hydroxycembra-1(15),2,11-trien-16,2-olide (2) [10] and $7\beta_8\beta$ -epoxy- 4β -hydroxycembra-1(15),2,11-trien-16,2-olide (3) [10], which have been isolated for the first time from a natural source. In addition, ten other compounds, including three further cembranoid derivatives (4–6), five sterols (7–11), and two open-chain compounds (12, 13) were isolated and identified.

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Results and Discussion. – Compound **1**, obtained as colorless crystals, showed the $[M + NH_4]^+$ signal at m/z 366.2268 (calc. 366.2280) in the HR-ESI mass spectrum, suggesting the molecular formula $C_{20}H_{28}O_5$, with seven degrees of unsaturation. The structure of **1** was deduced to correspond to 2-hydroperoxysarcophine, as elucidated on the basis of extensive spectroscopic analysis and comparison with the data of sarcophine (**5**), an epoxy cembranoid diterpene isolated from several marine organisms [11][12]. The relative configuration of **1** was determined from a NOESY experiment.

The IR absorptions of **1** at 3353, 1767, 1669, and 1256 cm⁻¹ indicated the presence of OH (or OOH), γ -lactone, olefin, and epoxide functionalities. The ¹H-NMR spectrum

(*Table*)¹) clearly indicated the presence of two olefinic H-atoms at $\delta(H)$ 5.47 (*s*, 1 H) and 4.99 (*t*, *J*=7.5 Hz, 1 H), and of four Me signals at $\delta(H)$ 1.90 (*s*), 1.56 (*s*), 1.17 (*s*), and 1.54 (*s*). The ¹³C-NMR (DEPT) spectrum (*Table*) exhibited 20 signals: seven quaternary C-atoms, three CH, six CH₂, and four Me groups, suggesting a diterpenoid skeleton for **1**. Four ¹³C-NMR signals for the tertiary olefinic C-atoms at $\delta(C)$ 151.1 (*s*), 147.9 (*s*), 131.2 (*s*), and 124.7 (*s*), as well as two signals for CH olefinic resonances at $\delta(C)$ 127.5 (*d*) and 112.8 (*d*) indicated that the molecule contained three C=C bonds. A further tertiary C-atom signal resonated at $\delta(H)$ 170.2 (*s*), which, together with the IR signal at 1767 cm⁻¹, suggested a γ -lactone. The signals at $\delta(C)$ 63.0 (*d*), $\delta(C)$ 60.2 (*s*), and $\delta(H)$ 2.41–2.43 (*m*, 1 H) indicated a trisubstituted epoxide in **1**. These IR and NMR data accounted for three of the five O-atoms. Therefore, the remaining two O-atoms and the oxygenated C-atom signal at $\delta(C)$ 85.0 (*s*) implied that **1**, most likely, possessed a hydroperoxy group. A positive reaction in the starch test (KI/AcOH) corroborated, indeed, that **1** was a peroxide [13].

Table. ¹*H- and* ¹³*C-NMR Data of* **1**. At 400 and 100 MHz, resp., in CDCl₃. Assignments¹) were corroborated by ¹H,¹H-COSY, HMQC, HMBC, and NOESY experiments.

	$\delta(H)$	$\delta(C)$		$\delta(\mathrm{H})$	$\delta(C)$
C(1)	-	151.1 (s)	H–C(11)	4.99(t, J=7.5)	127.5 (d)
C(2)	-	85.0 (s)	C(12)	-	131.2 (s)
H–C(3)	5.47 (s)	112.8(d)	$H_{a} - C(13)$	2.01 - 2.05 (m)	34.4(t)
C(4)	-	147.9 (s)	$H_{b}-C(13)$	2.09 - 2.15(m)	
$H_a - C(5)$	1.06 - 1.13 (m)	37.2 (t)	$H_{a} - C(14)$	1.31 - 1.37 (m)	23.2(t)
$H_b - C(5)$	1.83 - 1.89(m)	_	$H_{b}-C(14)$	2.12 - 2.17 (m)	
$H_a - C(6)$	1.86 - 1.88 (m)	23.1 (t)	H-C(15)	-	124.7 (s)
$H_b - C(6)$	2.00-2.05(m)	_	H-C(16)	-	170.2 (s)
$H_{\beta}-C(7)$	2.41 - 2.43 (m)	63.0(d)	Me(17)	1.90(s)	9.0(q)
C(8)		60.2(s)	Me(18)	1.56(s)	24.3(q)
$H_a - C(9)$	2.17 - 2.21 (m)	37.8 (t)	Me(19)	1.17(s)	17.0(q)
$H_{b}-C(9)$	2.33 - 2.37 (m)	-	Me(20)	1.54(s)	14.8(q)
$H_{a}-C(10)$	2.56 - 2.64(m)	22.4(t)	2-OOH	8.78 (s)	-
$H_{b} - C(10)$	2.67 - 2.71 (m)	-			

From the above IR, HR-ESI-MS, and NMR spectroscopic data, compound **1** was identified as a congener of sarcophine (**5**), a cembranoid diterpene previously isolated from the marine soft coral *Sarcophyton glaucum* [11][12]. The only difference was that the H-atom in 2-position of **5** was replaced by a OOH group in **1**.

The olefinic H-atom resonating at $\delta(H)$ 5.47 (*s*, H–C(3)) correlated with C(3) at $\delta(C)$ 112.8 in the HMQC spectrum of **1**. This H-atom displayed a ²*J*-type HMBC coupling to the oxygenated quaternary C(2) at $\delta(C)$ 85.0 (*Figure*). In addition, H–C(3) displayed a ³*J*-type HMBC correlation with the quaternary, olefinic C(1) at $\delta(C)$ 151.1. The olefinic resonance at $\delta(H)$ 4.99 (*t*), which correlated with C(11) at $\delta(C)$ 127.5 in the HMQC spectrum, was assigned to H–C(11). This H-atoms displayed ³*J*-type

¹) Arbitrary atom numbering.

HMBC correlations with C(9) at δ (C) 37.8, C(13) at 34.4, and C(20) at 14.8, respectively. Further HMBC correlations were observed between H–C(17) and C(1), C(2), C(14), C(15), and C(16); between H–C(18) and C(2), C(3), and C(5); between H–C(19) and C(7), C(8), and C(9); between H–C(20) and C(11), C(12), and C(13); and between H–C(3) and C(1), C(2), C(4), and C(5), respectively (*Figure*).

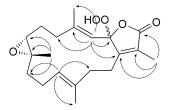


Figure. Selected HMBC correlations for 1

The relative configuration of **1** was established by a NOESY experiment. H-C(7) at $\delta(H) 2.41-2.43$ showed a NOESY correlation with Me(19) at $\delta(H) 1.17$, suggesting the same orientation for both groups. However, the configuration at C(2) remained unknown, since no cross-peak could be detected between the OOH H-atom and any other ¹H-NMR signal. From the above evidence, compound **1** was identified as 2-hydroperoxysarcophine. Although all experimental procedures, including extraction, isolation, and solvent evaporation, were conducted below 40°, it remains unclear whether **1** is a true natural product or an artifact.

Compounds 2 and 3 were obtained as colorless needles (m.p. 106-108 and $101-102^{\circ}$, resp.). Comparison of their NMR and MS spectroscopic data with those published showed that they correspond to 7β , 8β -epoxy- 4α -hydroxycembra-1(15),2,11-trien-16,2-olide and 7β , 8β -epoxy- 4β -hydroxycembra-1(15),2,11-trien-16,2-olide, respectively, which have previously been obtained by the bioconversion of sarcophine (5) [10], but, to the best of our knowledge, not isolated from a natural source.

Several other compounds were isolated and identified from *L. crassum*, including three further cembranoid diterpenes, sarcophytoxide (4) [14], sarcorphine (5) [11], and 14-hydroxycembra-1,3,7,11-tetraene (6) [15]; five sterols, including 4-methyl-ergo-stan-3-ol (7) [16], ergost-5-en-3-ol (8) [17], 17β ,20 β -epoxy-23,24-dimethylcholest-5-en-3 β ,22-diol (9) [18], pregnenolone (10) [19], and ergost-5-en-3,25-diol (11) [20]; as well as two other metabolites, hexadeca-7,11-dien-1-ol (12) [21] and hexadecanoic acid (13) [22].

As can be seen from the cembranoid molecular structures reported herein, all compounds contain an $\Delta^{1,15}$ rather than the known $\Delta^{15,17}$ unsaturation identified in previous constituents from *L. crassum*. To the best of our knowledge, this is the first report that *L. crassum* contains cembranoid diterpenes with both an OOH function and a $\Delta^{1,15}$ C=C bonds.

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Experimental Part

General. Column chromatography (CC) was performed with silica gel (200–300 mesh; Qingdao Haiyang Co., China), and anal. thin-layer chromatography (TLC) was performed with precoated silica gel GF_{254} plates (Qingdao Haiyang Co.). UV Spectra: Varian Cary-300-Bio spectrophotometer; λ_{max} (log ε) in nm. Optical rotation: Perkin-Elmer 341 polarimeter. IR Spectra: Nicolet Magna FT-IR-750 spectrometer; in cm⁻¹. ¹H- and ¹³C-NMR Spectra: Bruker AM-400 apparatus; at 400 (¹H) or 100 MHz (¹³C); chemical shifts δ in ppm rel. to residual CHCl₃ [δ (H) 7.26, δ (C) 77.0], coupling constants J in Hz. Assignments were supported by ¹H, ¹H-COSY, HMQC, and HMBC experiments. ESI- and HR-ESI-MS: Q-TOF Micro LC/MS/MS spectrometer; in m/z.

Animal Material. The soft coral Lobophytum crassum was collected in the sea waters of Hainan island in August 2004 from a depth of 8–10 m, and was identified by Prof. Z. C. Tang, Institute of Oceanology, Chinese Academy of Sciences, P. R. China, where a voucher specimen (No. 04B1) was deposited.

Extraction and Isolation. The whole specimen (1.5 kg dry weight after extraction) was extracted directly with EtOH at r.t. $(3 \times 7 \text{ d})$. The combined extracts were filtered and evaporated at reduced pressure ($<40^{\circ}$) to afford a crude residue (105 g), which was partitioned between AcOEt and H₂O, and between H₂O-sat. BuOH and H₂O, to afford an AcOEt and a BuOH-soluble fraction, resp. The AcOEt-soluble fraction (53 g) was purified by CC (SiO₂; AcOEt/petroleum-ether (PE) gradient): nine subfractions (*Fr.* 1–9) according to TLC. *Fr.* 2 (10 g) was further purified by repeated CC (SiO₂; AcOEt/PE 1:10 and AcOEt/CHCl₃ 1:30) to afford 6 (550 mg) and 12 (25 mg). *Fr.* 3 (4.5 g) was subjected to CC (SiO₂; AcOEt/PE 1:5 and AcOEt/CHCl₃ 1:15) to afford 1 (61 mg), 2 (10 mg), 3 (9 mg), 4 (300 mg), and 5 (200 mg). *Frs.* 4–6 were further separated by CC (SiO₂; acetone/PE 1:4 and AcOEt/CHCl₃ 1:2) to afford 7 (40 mg), 8 (3.5 g), 9 (460 mg), 13 (2 g), and a mixture of 10 and 11. The latter was further separated by CC (1. *Sephadex LH-20*, MeOH; 2. *RP-18*, MeOH/H₂O 9:1) to provide 10 (35 mg) and 11 (30 mg).

2-Hydroperoxysarcophine (=($1aR^{*},4E,11E,14aR^{*})$ -2,3,6,7,10a,13,14,14a-Octahydro-10a-hydroperoxy-1a,5,8,12-tetramethyloxireno[9,10]cyclotetradeca[1,2-b]furan-9(1aH)-one; **1**). Colorless crystals. M.p. 101–103°. UV (acetone): 212 (1.11). [a]²⁰_D = –153 (c=1.15, acetone). IR (KBr): 3353 (OOH), 2929, 1767 (γ -lactone), 1669 (olefin), 1448, 1385, 1326, 1256 (epoxide), 1064, 1033, 915, 732. ¹H- and ¹³C-NMR: see *Table*. EI-MS: 314 (7, [M – OOH+H]⁺), 297 (2), 271 (4), 231 (7), 205 (19), 191 (34), 166 (89), 124 (100). HR-ESI-MS: 366.2268 ([M + NH₄]⁺; C₂₀H₃₂NO⁺₅; calc. 366.2280).

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